

A Comparative Assessment of Biochemical Potential of Fresh and Fermented *Bamboosa balcooa* from Narengi (Guwahati), Assam

Paul Debalina¹, Das Baishali², Saikia Anamika², Gupta Abhijit^{1*} and Ahuja Vishal^{1,3}

1. University Institute of Biotechnology, Chandigarh University Mohali (Punjab), INDIA

2. Assam Downtown University, Panikhaiti, Guwahati – 781026, Assam, INDIA

3. University Centre for Research and Development, Chandigarh University Mohali (Punjab), INDIA

*abhijitgupta.biotech@cumail.in

Abstract

Traditional health practices and cultural heritage are filled with numerous underutilized plants having high healthcare potential that can prevent ailments. Bamboo is one such plant, distributed globally and is used in various cultural as well as food habits. Bamboo shoots have been consumed as fresh as well as fermented. The work was planned to assess the potential of both fresh and fermented bamboo shoots with their phytochemical profile. The fresh and fermented bamboo shoots were collected from Narengi (Guwahati), Assam and crushed for extract preparation. The extract was evaluated for respective antioxidant and antimicrobial potential along with the comparative study of the phytochemical profile.

The extracts have shown commendable antioxidant properties with IC_{50} 31.62 ± 1.67 mg.mL⁻¹ for fresh and 18.55 ± 2.78 mg.mL⁻¹ for fermented bamboo shoot extract. The extracts were also effective in inhibiting the growth of bacterial pathogens, especially *S. aureus*. Phytochemical analysis by GCMS identified the dominance of alcohols in fermented samples along with salicylic acid derivatives which may be crucial for bioactivities. Bamboo shoots have shown positive results in terms of healthcare applications. However, still there is a lot of space to fill to make a commercial formulation to cure disease or malnutrition.

Keywords: Fermented shoot extract, phytochemical analysis, antioxidants, antimicrobial.

Introduction

Bamboos, grass of the Poaceae family with more than 148 species under 29 genera are distributed all over the world and are spread over an area of 9.57 million hectares all over India²⁶. Bamboo is mostly found in tropical, subtropical, semi-evergreen, moist deciduous areas of forest. It is also referred to as poor man's timber and rich man's delicacy due to its widespread availability and multifaceted applications². It plays a crucial role in the prevention of soil erosion and conservation of soil moisture. Bamboo shoots are rich in proteins, carbohydrates, vitamins, fibers and minerals and are very low in fat¹⁰, hence shoots or sprouts are edible and are used as a traditional delicacy mostly in Southeast Asian

and East Asian countries² including China, Japan, the US, North East India, Thailand, Nepal, Bhutan, Korea, Australia, New Zealand, Malaysia and Indonesia²⁴.

Fresh bamboo shoots also contain taxiphyllin, cyanogenic glycoside that is toxic and bitter. Bamboo shoots of all the species are not edible²². The most commonly edible bamboo shoots in India are *Bambusa pallid*, *Bambusa tulda*, *Bambusa polymorpha*, *Bambusa balcooa*, *Dendrocalamus giganteus*, *Dendrocalamus hamiltoni* and *Melocanna bambusoides*¹¹.

Bamboo shoots exhibit great potential as a nutritious food resource, thus most commonly consumed as well as used as stored reserve food in North Eastern India^{7,23}. Mostly, bamboo shoots are consumed in raw, dried, canned, boiled and fermented forms, depending upon the traditional and local culture and food habits^{10,28}. It is considered nutritious and Rich in antioxidant compounds and vitamins including thiamine, niacin, vitamin A, vitamin B6, vitamin E, minerals and electrolytes including iron, calcium and phosphorus that can potentially be helpful in preventing metabolic disorders¹⁵. Besides raw and dried form consumption, bamboo shoots are also fermented before consumption to extend shelf life and enhance the flavour²⁸. In addition, bamboo shoots also contain unsaturated fats that help in fighting against bad cholesterol⁴.

Available literature has proven the health benefits including antioxidant, anti-aging and anti-cancer properties, cardiovascular disease prevention, weight loss, improved digestion, decreased blood pressure and antimicrobial activity due to flavones and glycosides^{1,14,23}. However, their consumption is currently limited to tribal people in India and there is a need to create awareness among the general public about their nutritional benefits. The proposed work was planned to evaluate the potential of fresh and fermented bamboo shoots.

Material and Methods

Plant sample collection: The fresh sample, *Bambusa balcooa* shoots and fermented bamboo shoots were collected from Narengi, Guwahati, Assam in September 2021 (Fig. 1). The fresh bamboo shoots were cleaned using tap water and were weighed in the electronic balance. After weighing, both fresh bamboo shoots and fermented bamboo shoots were crushed using mortar and pestle and were also weighed.

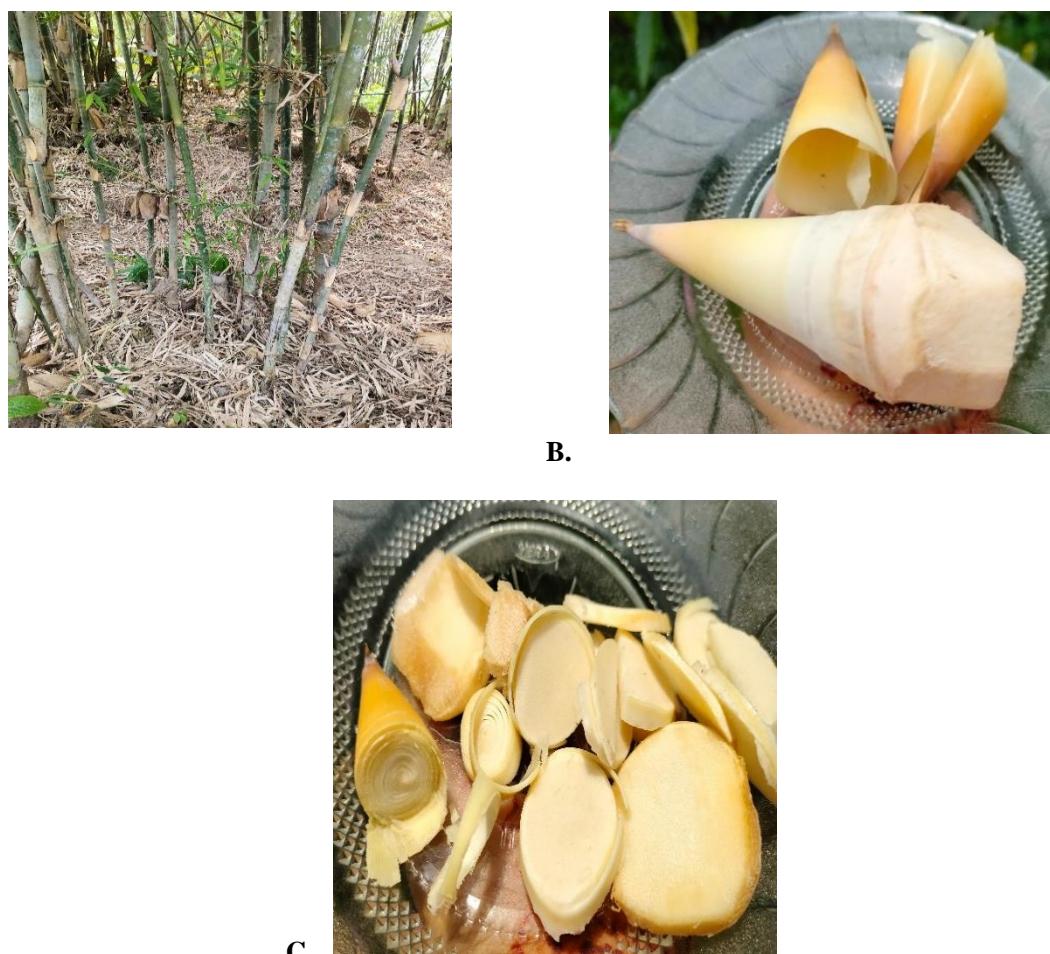


Fig. 1: A. *Bamboosa balcooa*, B. collected sample of *Bamboosa balcooa* Shoot and C. fresh *Bambusa balcooa* Shoot

Preparation of plant extract: The shoot extract was prepared by solvent-assisted extraction as defined earlier¹¹. 40 g of both fresh and fermented crushed plant was added in six beakers and was dipped in 60 ml of methanol, ethanol and acetone respectively. The soaked samples were kept for 72 hours to ensure complete extraction. The extracts were then filtered using filter paper for 24 hours. After the filtration process, the filtrates were transferred into six test tubes respectively and were covered in parafilm and aluminium foil and stored in the refrigerator.

Antioxidant potential: Antioxidant substances and mechanisms are required to protect cells against free radicals and oxidative stress and subsequently play a vital role in lowering the risk of cardiac ailments, cancer and other associated diseases. Measuring the antioxidant activity of bioactive formulations is therefore essential to ensure the quality of functional foods. The antioxidants potential of fresh and fermented bamboo shoot extract was assessed by DPPH, ABTS, FRAP and CUPRAC methods against different types of free radicle species.

For the DPPH assay, fresh and fermented shoot extracts were screened against 1,1-diphenyl-2-picryl hydrazyl (DPPH). The stock of 0.1 μ g/10 μ l was used for assessment in different concentrations. 10-1000 μ l stock solution was mixed separately with 3 mL DPPH workable solutions, mixed and

kept at room temperature for 30 min in the dark. The absorbance of solutions was recorded at 517 nm⁵. A similar strategy was also adopted for ABTS assay¹⁹, FRAP assay¹³ and CUPRAC assay³. The antioxidant potential was calculated in comparison to 100 μ l of 1mg.mL⁻¹ TROLOX, as the positive control and distilled water as a negative control. The antioxidant potential was calculated using eq. 1:

$$\text{Antioxidant potential (\%)} = \frac{\text{Abs}_{\text{control}} - \text{Abs}_{\text{Sample}}}{\text{Abs}_{\text{Control}}} \times 100$$

Antimicrobial potential: The antimicrobial potential of extract of fresh and fermented bamboo shoots was determined with different concentrations of extract against clinical microbial pathogens by CFU count^{6,9}. Test bacterial pathogens were revived on nutrient broth and incubated at 37°C to attain a microbial load of 1.5 \times 10⁸ CFU.ml⁻¹. Shoot extracts of 1mg.10ml⁻¹ stock, prepared in dimethyl sulfoxide were used for evaluation. Different concentrations of sheet extract were added to each well and incubated at 37°C. 1mg.ml⁻¹ of antimicrobial was used as positive control while DMSO was used as negative control. The plates were inoculated for 24-36 hours at 37°C (bacteria) and 27°C (fungi). Inhibition (%) was assessed by CFU and compared with negative control (100%).

Comparative analysis of phytochemicals in fresh and fermented bamboo shoot: The plant extract was characterized by different phytochemicals including tannins, phenolics, proteins, lipids and carbohydrates. Total phenolic content in plant extract was determined by Folin-Ciocalteu (FC) reagent as defined by Singleton and Rossi²⁹. 100µL of plant extract was mixed with 500µL diluted FC reagent solution and kept at 22°C for 5 min. Post incubation, 2 mL Na₂CO₃ (20% aqueous solution) was added to the solution and further incubated at 22°C for 90 min. The absorbance was recorded at 650 nm and phenolics was quantified with gallic acid standard (1mg.mL⁻¹). Similarly, total flavonoid content was quantified by using aluminum chloride (AlCl₃) method²⁷. For quantification, quercetin was used as a standard and absorbance was recorded at 510 nm. The total tannin content of plant extract was determined by titrimetric method as defined in Ayurvedic Pharmacopoeia of India (<https://www.ayurveda.hu/api/API-Vol-1.pdf>).

The total saponin content in plant extract was determined with the vanillin-sulfuric acid colorimetric method. The absorbance of the sample was determined at 544 nm and

quantified with diosgenin equivalent (DE).g⁻¹¹⁶. Total and reducing sugar content and protein content were quantified by phenol sulfuric²⁰, dinitro-salicylic acid²¹ and Bradford method⁸.

Statistical analysis: The statistical analysis of antimicrobial and antioxidant analysis was done by using MS-Excel with a data analysis pack using ANOVA. The p-value was considered for model suitability which must be below 0.05 for model fitness.

Results and Discussion

Bamboo shoot dried powder was extracted with hydro-alcohol (1:1) for 72 hours at room temperature and filtered to remove solid fraction. The extract was characterized for phytochemical constituents like phenolics, carbohydrates and other metabolites. The bioactivities of both fresh and fermented bamboo shoots were assessed for antioxidants and antimicrobial activity. The results pertaining to various analyses have been summarised.

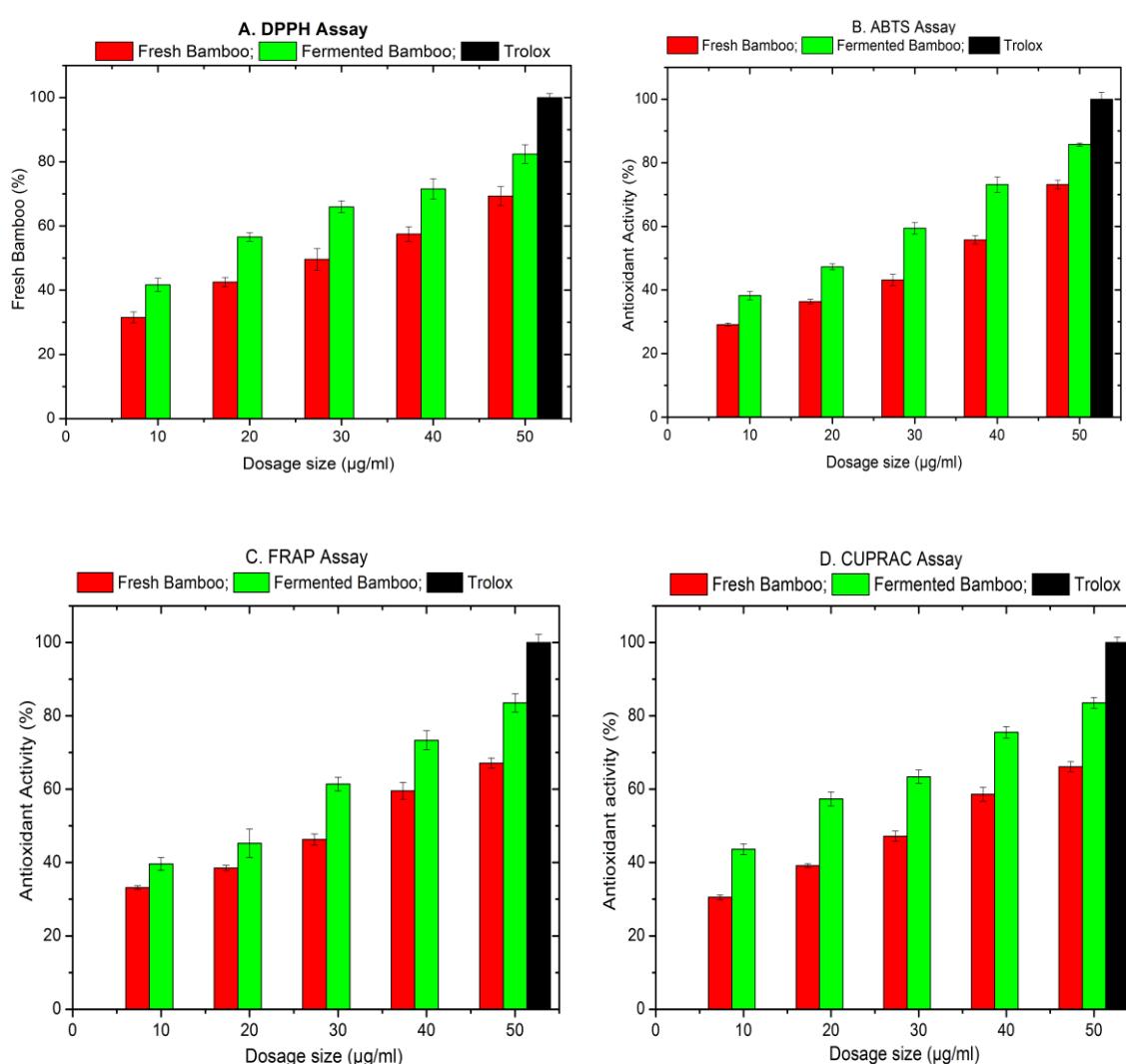


Fig. 2: Antioxidant potential of fresh and fermented bamboo shoot extract by A. DPPH assay, B. ABTS assay, C. FRAP assay and D. CUPRAC assay.

Antioxidants: The antioxidant potential of plant extract was determined against 2,2-diphenyl-1-picrylhydrazyl radical (DPPH), 2,2'-azino-bis(3-ethyl benzothiazoline-6-sulfonic acid) diammmonium radical cation (ABTS), Fe^{3+} -TPTZ (FRAP) and $\text{Cu}(\text{Nc})_2^{2+}$ (CUPRAC). The test involved different types of radicals including organic and metal-based structures to determine the total antioxidant potential. For comparison, trolox was used as a positive control (commercial antioxidant compound) and distilled water as a negative control.

As depicted in fig. 2, both fresh and fermented bamboo shoot extract have shown antioxidant potential in a dose-dependent manner in all four assays i.e. DPPH (Fig. 2A), ABTS (Fig. 2B), FRAP (Fig. 2C) and CUPRAC (Fig. 2D). However, the antioxidant potential of fermented bamboo shoot was higher than fresh bamboo shoot extract. The maximum antioxidant potential of shoot extract was exhibited at a higher concentration range of $50\text{ }\mu\text{g.mL}^{-1}$. The antioxidant potential was still high in comparison to plant extract. Bamboo shoots have been identified as a rich source of antioxidant compounds including phenols, flavonoids, vitamins and minerals which contribute significantly to its antioxidant activity²².

Iwansyah et al¹⁸ reported that the ethanolic extract of bamboo shoots contained a high amount of $27.83\text{ mg GAE.g}^{-1}$ phenolics and 2.49 mg QE.g^{-1} flavonoids. The IC_{50} of ethanolic extract was $347.48\text{ }\mu\text{g.mL}^{-1}$ for the Kuning bamboo shoot. Santosh et al²⁵ reported IC_{50} of fortified bamboo shoot $2100.95\text{ }\mu\text{g.L}^{-1}$. IC_{50} values of bamboo shoot extract in current work were $18.55\pm2.78\text{ }\mu\text{g.mL}^{-1}$ (fermented shoot) and $31.62\pm1.67\text{ }\mu\text{g.mL}^{-1}$ (fresh shoot). The values were quite lower in comparison to previous work and clearly visible that fermented shoot extract has higher antioxidant potential.

Antimicrobial analysis: Fresh as well as fermented bamboo shoot extracts have shown a wide range of phytochemicals

that are mainly responsible for antimicrobial activity. The antimicrobial activity of extracts has shown a similar trend as fermented bamboo shoot extract has higher inhibitory potential than fresh bamboo shoots (Fig. 3). Both fresh and fermented bamboo shoot extracts were effective against microbial pathogens and inhibited the growth of bacterial as well as fungal pathogens. The Antimicrobial activity was maximum against *S. aureus* with $75.39\pm1.92\text{ %}$ (fermented bamboo shoot) and $67.51\pm1.11\text{ %}$ (fresh bamboo shoot) while minimum against *Aspergillus niger* with $17.83\pm0.35\text{ %}$ (Fresh bamboo shoot) and $19.89\pm0.76\text{ %}$ (fermented bamboo shoot).

The results from the current work coincide with the previous researches. Ethanolic extract of bamboo shoot has antimicrobial activity against salivary *Streptococcus mutans* and *Lactobacillus acidophilus* with inhibition zones of $23.00\pm0.816\text{ mm}$ and $22.00\pm0.816\text{ mm}$. The maximum inhibition was reported at a bamboo shoot extract concentration of $60\text{ }\mu\text{g.mL}^{-1}$ ¹². In another work, fermented shoot extract has shown antimicrobial activity against *S. aureus*, *P. aeruginosa* and *E. coli*³⁰. The antimicrobial activity of bamboo shoots was possibly due to chitin-binding proteins (Pp-AMP1 and Pp-AMP2), flavones and glucoside¹².

Statistical analysis: The antioxidant and antimicrobial analysis were evaluated for fitness by two-way ANOVA. The 'p' value must be below 0.05 to make a model fit. Regarding antioxidant analysis, the p-values for DPPH, ABTS, FRAP and CUPRAC analysis were well below 0.05. For DPPH the p values were 0.00005 and 0.03, for ABTS, 0.00004 and 0.01, for FRAP, 0.00005 and 0.01 and CUPRAC, 0.00005 and 0.03. The ANOVA analysis supported the model's fitness. Similar kind of observations was reported with antimicrobial analysis. The p-value for antimicrobial analysis was 0.039 which was also below 0.05 (Table 1).

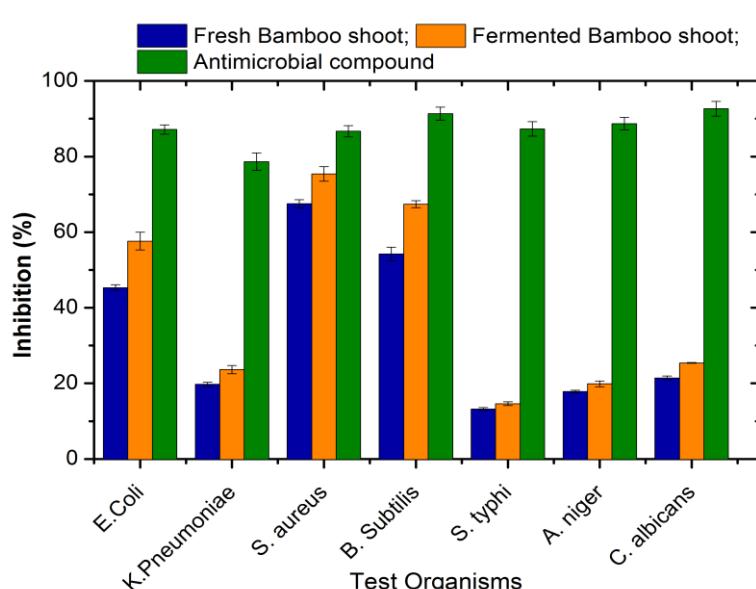


Fig. 3: Antimicrobial activity of fresh and fermented bamboo shoot

Table 1
ANOVA analysis of antioxidant and antimicrobial activity of bamboo plant extract

Source of Variation	SS	Df	MS	F	P-value	F crit
DPPH Assay						
Rows	0.55	2	0.27	43.12	5.19E-05	4.46
Columns	0.13	4	0.03	5.04	0.03	3.84
ABTS Assay						
Rows	0.62	2	0.31	44.65	4.57E-05	4.46
Columns	0.19	4	0.05	6.89	0.01	3.84
FRAP Assay						
Rows	0.53	2	0.27	43.74	4.93E-05	4.46
Columns	0.17	4	0.04	7.15	0.01	3.84
CUPRAC Assay						
Rows	0.58	2	0.29	44.64	4.58E-05	4.46
Columns	0.13	4	0.03	5.02	0.03	3.84
Antimicrobial assessment						
Rows	1740.93	6	290.16	2.47	0.039	2.32
Columns	81481.78	7	11640.25	98.95	5.13E-24	2.24

Table 2
Phytochemical analysis of fresh and fermented bamboo shoot extract

Phytochemicals	Bamboo shoot extract (g.100g ⁻¹)		Change (Rise/reduction)
	Fresh	Fermented	
Carbohydrate	9.63±0.23	1.11±0.01	-88.47
Proteins	2.89±0.11	5.83±0.2	101.73
Phenolics	3.1±0.19	2.9±0.01	-6.45
Flavonoid	0.3 ±0.02 (ruetin equivalent)	0.34±0 (ruetin equivalent)	13.33
Tannins	0.03±0	0.03±0	0.0
Saponins	0.15±0.0	0.14±0.03	-6.67
Total sugars	4.51±0.19	2.91±0.17	-35.48

- Reduction and + Increase

Comparative analysis of phytochemical profiling: The phytochemicals are the root factor for bioactivities. Both fresh as well as fermented bamboo shoot extracts were characterized for the presence of various phytochemicals including carbohydrates, proteins, tannin, flavonoids, phenolics etc. (Table 2). Fresh shoot extract contains a significant amount of saponins, phenolics and flavonoids. However, the concentration of different phytochemicals has changed significantly in fermented shoot extract due to microbial action. Carbohydrate and sugar content have been reduced by more than 88% and 35% respectively as it is the main substrate for microbial fermentation while protein content was increased by more than 101% due to the additional production of proteins by microorganisms in fermentation.

Literature has also reported the presence of diverse phytochemicals including phenols, derivatives, acids, alcohols and flavonoids. Iwansyah et al¹⁸ have compared different types of bamboo shoots like Kuning, Gombong, Betung and Buluh among which maximum phenolics and flavonoids were reported from Buluh and Betung respectively. The maximum antioxidant potential was

reported from Betung shoots ethanolic extract. Comparative GCMS profiles of both fresh and fermented shoot extract also revealed the dominance of alcohols³¹. The bamboo shoot extracts have also shown the hepatoprotective effects of fresh and fermented bamboo shoot extract in diabetic mice models.

In comparison to fresh, fermented shoot extracts have shown a better reduction in liver enzyme levels including serum glutamic pyruvic transaminase, serum glutamic-oxaloacetic transaminase, alkaline phosphatase and bilirubin¹⁷. Some of the recent studies have also reported the valuable contribution of plant-based products and phytochemicals in health care and in improving food nutrition³²⁻³⁴. The difference in activity is mainly attributed to diverse phytochemicals.

In addition, bamboo shoots also contain a good amount of vitamins and minerals that become a conducive factor for bioactivities. The results seem promising but still, cell line-based and animal model-based analysis are required to conclude its commercial feasibility as health care products.

Conclusion

Phytochemicals including primary and secondary metabolites have shown a diverse range of bioactivity and have shown a brighter side of healthcare and herbal formulations. Fresh and fermented bamboo shoot extracts have shown the presence of phenolics, alcohols, acids, flavonoids, tannins and proteins which are mainly responsible for bioactivities including antioxidants and antimicrobial, anti-aging and anti-fatigue. The fermented bamboo shoot extract has a significantly higher potential than fresh.

References

1. Aghababaei F. and Hadidi M., Recent Advances in Potential Health Benefits of Quercetin, *Pharmaceuticals*, **16**(7), 1020, <https://doi.org/10.3390/PH16071020> (2023)
2. Ahmad Z., Upadhyay A., Ding Y., Emamverdian A. and Shahzad A., Bamboo: Origin, Habitat, Distributions and Global Prospective, *Biotechnological Advances in Bamboo: The “Green Gold” on the Earth*, Springer, 1–31, https://doi.org/10.1007/978-981-16-1310-4_1 (2021)
3. Apak R., Güçlü K., Özyürek M. and Karademir S.E., Novel Total Antioxidant Capacity Index for Dietary Polyphenols and Vitamins C and E, Using Their Cupric Ion Reducing Capability in the Presence of Neocuproine: CUPRAC Method, *J. Agric. Food Chem.*, **52**(26), 7970–7981, <https://doi.org/10.1021/JF048741X> (2004)
4. Ashour A., Elbermawi A., Amen Y., Allam A.E., Ikeda H., Nagata M., Kumagae K., Azuma T., Taguchi A., Takemoto T., Matsumoto M. and Shimizu K., Melanin Synthesis Inhibition Activity of Compounds Isolated from Bamboo Shoot Skin (*Phyllostachys pubescens*), *Molecules*, **28**(1), 23, <https://doi.org/10.3390/molecules28010023> (2022)
5. Baliyan S., Mukherjee R., Priyadarshini A., Vibhuti A., Gupta A., Pandey R.P. and Chang C.M., Determination of Antioxidants by DPPH Radical Scavenging Activity and Quantitative Phytochemical Analysis of *Ficus religiosa*, *Molecules*, **27**(4), 1326, <https://doi.org/10.3390/molecules27041326> (2022)
6. Beal J. et al, Robust estimation of bacterial cell count from optical density, *Commun. Biol.*, **3**(1), 1–29, <https://doi.org/10.1038/s42003-020-01127-5> (2020)
7. Bisht M.S., Nirmala C. and Meetei O.S., Bamboo shoots for food in North-East India: conventional and contemporary, Proceedings of 10th World Bamboo Congress, Damyang, 17 (2015)
8. Bradford M.M., A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding, *Anal. Biochem.*, **72**, 248–254, [https://doi.org/10.1016/0003-2697\(76\)90527-3](https://doi.org/10.1016/0003-2697(76)90527-3) (1976)
9. Campbell J., High-throughput assessment of bacterial growth inhibition by optical density measurements, *Curr. Protoc. Chem. Biol.*, **3**(3), 195–208 (2011)
10. Chongtham N. and Bisht M.S., Bamboo Shoot: Superfood for Nutrition, Health and Medicine, *Bamboo Shoot*, CRC Press Boca Raton, <https://doi.org/10.1201/9781003032939> (2020)
11. Choudhury D., Sahu J.K. and Sharma G.D., Value addition to bamboo shoots: a review, *J. Food Sci. Technol.*, **49**(4), 407, <https://doi.org/10.1007/S13197-011-0379-Z> (2012)
12. Divya Bharathi S., Priya Deepa Lakshmi K., Gunasekaran M., Venkata Lakshmi S., Reji A. and Kathija Sulthana F., Comparison of the Antibacterial Efficacy of Bamboo Shoot Ethanol Extract with Chlorhexidine Mouth Rinse Against Salivary *Streptococcus mutans* and *Lactobacillus acidophilus*: An Ex Vivo Study, *Cureus*, **16**(1), e53085, <https://doi.org/10.7759/cureus.53085> (2024)
13. Fernandes R.P.P., Trindade M.A., Tonin F.G., Lima C.G., Pugine S.M.P., Munekata P.E.S., Lorenzo J.M. and de Melo M.P., Evaluation of antioxidant capacity of 13 plant extracts by three different methods: cluster analyses applied for selection of the natural extracts with higher antioxidant capacity to replace synthetic antioxidant in lamb burgers, *J. Food Sci. Tech.*, **53**(1), 451–460, DOI: 10.1007/s13197-015-1994-x (2016)
14. Fusi F., Trezza A., Tramaglino M., Sgaragli G., Saponara S. and Spiga O., The beneficial health effects of flavonoids on the cardiovascular system: Focus on K⁺ channels, *Pharmacol. Res.*, **152**, 104625, <https://doi.org/10.1016/J.PHRS.2019.104625> (2020)
15. Harenčář L., Ražná K. and Nôžková J., Cyanogenic Glycosides - Their Role and Potential In Plant Food Resources, *J. Microbiol. Biotechnol. Food Sci.*, **11**(3), e4771–e4771, <https://doi.org/10.15414/JMBFS.4771> (2021)
16. Hiai S., Oura H. and Nakajima T., Color reaction of some saponins and saponins with vanillin and sulfuric acid, *Planta Med.*, **29**(2), 116–122, <https://doi.org/10.1055/s-0028-1097639> (1976)
17. Indira A., Joshi B., Koul A. and Chongtham N., Comparative hepato-ameliorative effects of *Bambusa nutans* fresh and fermented shoot extracts on STZ induced diabetic LACA mice, *Talanta*, **274**, 126035, <https://doi.org/10.1016/J.TALANTA.2024.126035> (2024)
18. Iwansyah A.C., Kumalasari R., Darmajana D.A. and Ratnawati L., Antioxidant properties and toxicity of selected bamboo shoots “iwung” extract: a comparative study, IOP Conference Series: Earth and Environmental Science, **251**(1), 012017, <https://doi.org/10.1088/1755-1315/251/1/012017> (2019)
19. Lee K.J., Oh Y.C., Cho W.K. and Ma J.Y., Antioxidant and Anti-Inflammatory Activity Determination of One Hundred Kinds of Pure Chemical Compounds Using Offline and Online Screening HPLC Assay, *Evid. Based Complement. Alternat. Med.*, <https://doi.org/10.1155/2015/165457> (2015)
20. Masuko T., Minami A., Iwasaki N., Majima T., Nishimura S.I. and Lee Y.C., Carbohydrate analysis by a phenol–sulfuric acid method in microplate format, *Anal. Biochem.*, **339**(1), 69–72, <https://doi.org/10.1016/J.AB.2004.12.001> (2005)
21. Miller G.L., Use of Dinitrosalicylic Acid Reagent for Determination of Reducing Sugar, *Anal. Chem.*, **31**(3), 426–428, <https://doi.org/10.1021/ac60147a030> (1959)
22. Nirmala C., Bisht M.S., Bajwa H.K. and Santosh O., Bamboo: A rich source of natural antioxidants and its applications in the food and pharmaceutical industry, *Trends Food Sci. Technol.*, **77**, 91–99, <https://doi.org/10.1016/J.TIFS.2018.05.003> (2018)

23. Nongdam P. and Tikendra L., The Nutritional Facts of Bamboo Shoots and Their Usage as Important Traditional Foods of Northeast India, *Int. Sch. Res. Notices*, 1–17, <https://doi.org/10.1155/2014/679073> (2014)

24. Parveen H., Tewari L., Pradhan D. and Chaudhary P., Comparative Study of Diverse Pretreatment Approaches to Degrade Lignin from Bambusa balcooa, *Bio Resources*, **17**(4), 5578–5599, DOI: 10.15376/biores.17.4.5578-5599 (2022)

25. Santosh O., Bajwa H.K., Bisht M.S. and Chongtham N., Antioxidant activity and sensory evaluation of crispy salted snacks fortified with bamboo shoot rich in bioactive compounds, *Appl. Food Res.*, **1**(2), 100018, <https://doi.org/10.1016/j.afres.2021.100018> (2021)

26. Sharma M. and Nirmala C., Bamboo Diversity of India: An update, 10th World Bamboo Congress, Korea, 17–22 (2015)

27. Shraim A.M., Ahmed T.A., Rahman M.M. and Hijji Y.M., Determination of total flavonoid content by aluminum chloride assay: A critical evaluation, *LWT*, **150**, 111932, <https://doi.org/10.1016/J.LWT.2021.111932> (2021)

28. Singhal P., Satya S. and Naik S.N., Fermented bamboo shoots: A complete nutritional, anti-nutritional and antioxidant profile of the sustainable and functional food to food security, *Food Chem.: Mol. Sci.*, **3**, 100041, <https://doi.org/10.1016/J.FOCHMS.2021.100041> (2021)

29. Singleton V.L. and Rossi J.A., Colorimetry of Total Phenolics with Phosphomolybdic-Phosphotungstic Acid Reagents, *Am. J. Enol. Vitic.*, **16**(3), 144–158, DOI: 10.5344/ajev.1965.16.3.144 (1965)

30. Sonia Angeline M., Challaraj Emmanuel E.S., Nonglait R. and Suting B., Antimicrobial and Antioxidant activity of Fermented Bamboo Shoot Dendrocalamus hamiltonii, *Curr. Trends Biotechnol. Pharm.*, **15**(5), 425–436, <https://doi.org/10.5530/CTBP.2021.3S.36> (2021)

31. Zheng J., Zhang F., Zhou C., Lin M. and Kan J., Comparison of Flavor Compounds in Fresh and Pickled Bamboo Shoots by GC-MS and GC-Olfactometry, *Food Sci. Technol. Res.*, **20**(1), 129–138, <https://doi.org/10.3136/FSTR.20.129> (2014).

(Received 23rd July 2024, accepted 25th September 2024)